



ALLIANCE™

(<https://www.globalseafood.org>).



Health &
Welfare

Establishing nitrifying bacteria in super-intensive biofloc shrimp production

1 May 2011

By Clete A. Otoshi, Neil Rodriguez and Shaun M. Moss, Ph.D.

Bacterial community useful in start-up of a system, after disinfection following a disease outbreak



Green microalgae typically persist in the water columns of super-intensive shrimp trials conducted at Oceanic Institute.

The use of biofloc technology for super-intensive shrimp production is emerging as a sustainable alternative to traditional shrimp culture. The advantages of super-intensive biofloc production include temperature control for year-round production, the ability to minimize the introduction and spread of shrimp pathogens, and drastic reductions in water use and effluent discharge. Biofloc management relies on in situ microbes, including bacteria and algae, to prevent the accumulation of the inorganic nitrogenous metabolites ammonia and nitrite, which can be toxic to shrimp.

A heterotrophic bacterial community can minimize ammonia accumulation through assimilation into bacterial biomass. This bacterial community can be promoted by manipulating carbon:nitrogen ratios in the water column through the use of low-protein feeds and the addition of an organic carbon source such as molasses.

A chemoautotrophic bacterial community (i.e., nitrifying bacteria) can prevent the accumulation of ammonia and nitrite through oxidation to nitrate. This bacterial community can be maintained by replacing alkalinity consumed during nitrification with an inorganic carbon source such as sodium bicarbonate or calcium carbonate.

Depending upon light availability and management strategy, photoautotrophs (i.e., microalgae) can also be present. They minimize ammonia and nitrate accumulation through assimilation into algal biomass.



A comprehensive solution for the wild seafood supply chain.

- ✓ Crew rights
- ✓ Food safety
- ✓ Environmental responsibility

Best Seafood Practices

LEARN MORE >

(<https://bspcertification.org/>).

Heterotrophic bacteria have short generation times and can be established quickly, whereas nitrifying bacteria grow and divide more slowly and require careful management in a biofloc system. Biofloc strategies for super-intensive shrimp production typically include members of all three of these microbial communities.

Establishing nitrifying bacteria

Experienced home aquarists understand that “aged” biofilter media or gravel, which has an established nitrifying bacterial community, is helpful when starting a new aquarium. If such media are not available, the aquarium can be allowed to “cycle” naturally by maintaining an appropriate number and type of fish until cycling is completed.

Fish used during cycling excrete ammonia, which serves as a substrate for ammonia-oxidizing bacteria (AOB). Ammonia is oxidized by the bacteria to nitrite, which in turn provides substrate for nitrite-oxidizing bacteria (NOB). Nitrite is oxidized by NOB to relatively non-toxic nitrate. In these steps, ammonia or nitrite must accumulate in the water before the respective nitrifying bacterial groups become established.

This cycling process, which can take four to six weeks, is characterized by spikes in ammonia and nitrite concentrations as AOB and NOB become established. Several strategies can accelerate the cycling process and minimize the spikes.

One such strategy is to maintain a minimal level of ammonia and/or nitrite at the start of the cycle through the chemical addition of ammonium chloride and/or sodium nitrite. This provides substrate for nitrifying bacteria immediately, rather than waiting for ammonia and nitrite to accumulate naturally. If cycling is not conducted properly in a home aquarium, “new tank syndrome,” where ammonia and/or nitrite can increase to lethal levels, can result.

The same concepts apply to the start-up of biofloc systems used for super-intensive shrimp production. Importantly, issues associated with improper cycling are exacerbated by super-intensive stocking densities and rapid shrimp growth. Feed rates and subsequent ammonia and nitrite production are high, and with insufficient cycling, this can result in much faster and higher spikes of ammonia and/or nitrite that can have sub-lethal or lethal effects on the shrimp.



Super-intensive shrimp trials were conducted in Oceanic Institute's biofloc raceway system.

Super-intensive biofloc system

Research on super-intensive shrimp culture with biofloc has been conducted at Oceanic Institute in Waimanalo, Hawaii, USA, since 1997. The institute maintains a selective-breeding program for Pacific white shrimp, *Litopenaeus vannamei*, which requires growout evaluation of selected families. These trials are conducted in a 75-cubic-meter super-intensive BFT raceway stocked at 300-400 shrimp per cubic meter in Oceanic Institute's Nucleus Breeding Center.

Strict biosecurity protocols in the breeding center protect the valuable stocks from potential pathogens. The raceways are chlorinated and rinsed with freshwater between trials. Each new super-intensive trial is started with clear seawater.

Difficulties in establishing adequate nitrifying bacterial communities during the cycling periods of several past trials resulted in nitrite spikes that sometimes exceeded 25 mg/L nitrite-nitrogen. Such elevated nitrite concentrations resulted in poor shrimp health and at times has required flushing of the system.

Better control of the spikes in nitrite during cycling was needed. Techniques such as the addition of biofilter backwash, nursery system water and starter nitrifying bacterial cultures resulted in limited success. In addition, these resources were not consistent in quality and availability, or were costly.

Sodium nitrite addition

Nitrite-oxidizing bacteria have a doubling time of at least 15 hours, whereas ammonia-oxidizing bacteria can double in about half that time. The shorter generation time likely explains the observation that ammonia cycling spikes in Oceanic Institute's super-intensive biofloc system never reached toxic

levels, and AOB became established more easily than NOB.

Because nitrite toxicity was a primary concern, experiments with the addition of nitrite in the form of sodium nitrite were conducted. The addition of sodium nitrite prior to stocking shrimp would provide the substrate necessary to allow naturally occurring NOB to become established immediately. Importantly, nitrifying bacteria are sensitive to ultraviolet radiation when unattached, so black shade cloths were installed over the raceway during cycling.

Fig. 1 shows nitrite-nitrogen concentrations in two trials in which multiple doses of sodium nitrite were added (trials 4 and 5) and prior trials where nothing was added (trial 1). In trial 2, biofilter backflush and one dose of sodium nitrite were added. Only biofilter backflush was added in trial 3.

Fig. 1: Nitrite-nitrogen concentrations during the cycling period of super-intensive shrimp trials.

During trial 5, the biofloc raceway was filled two weeks prior to stocking and initially spiked with 1-2 mg/L of sodium nitrite-nitrogen. Five subsequent doses were added to the raceway in an attempt to maintain a minimal nitrite concentration. Fig. 2 shows total ammonia-nitrogen, nitrite-nitrogen and nitrate-nitrogen prior to stocking and during the cycling period of trial 5, which was stocked at a density of 300 shrimp per cubic meter.

Fig. 2: Trial 5 ammonia-nitrogen, nitrite-nitrogen and nitrate-nitrogen concentrations two weeks prior to and during the cycling period. Red asterisks indicate sodium nitrite additions.

The increase in nitrate-nitrogen just days after sodium nitrite was first added confirmed the establishment of NOB by demonstrating that nitrite was being oxidized to nitrate. The presence of NOB early in the cycling period presumably allowed this group of bacteria to react more quickly to the natural accumulation of nitrite after shrimp were stocked.

No significant spikes in nitrite concentration were observed during or subsequent to the cycling period of trial 5. Importantly, expensive nitrifying bacteria starter cultures were not needed, as there were sufficient nitrifiers in the incoming seawater to develop a community rapidly when provided with an appropriate substrate.

Perspectives

There are many scenarios where a simple, cost-effective protocol to establish a nitrifying bacterial community would be useful, such as initial start-up of a system, start-up after disinfection following a disease outbreak, lack of aged water, the desire to limit cross-contamination with aged water or the start-up of intermittent research trials.

During trial 5, 4.3 kg of sodium nitrite were used during the cycling period for the 75-cubic-meter raceway at a cost of \$10.63. Data from trial 5 suggested that a shorter start-up period and less sodium nitrite dosing may be adequate to complete proper cycling. Future investigations will focus on optimizing the timing and amount of sodium nitrite dosage in an effort to further reduce cost and labor.

(Editor's Note: This article was originally published in the May/June 2011 print edition of the Global Aquaculture Advocate.)

Authors



CLETE A. OTOSHI

Oceanic Institute
41-202 Kalanianaʻole Highway
Waimanalo, Hawaii 96795 USA

cotoshi@oceanicinstitute.org (<mailto:cotoshi@oceanicinstitute.org>).



NEIL RODRIGUEZ

Oceanic Institute
41-202 Kalanianaʻole Highway
Waimanalo, Hawaii 96795 USA



SHAUN M. MOSS, PH.D.

Oceanic Institute
41-202 Kalanianaʻole Highway
Waimanalo, Hawaii 96795 USA

Copyright © 2024 Global Seafood Alliance

All rights reserved.